Coconut water alters maternal high fat diet induced changes in hormones and pup morphometry of Wistar rats

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Abstract

Background: Maternal high fat diet (HFD) during gestation adversely programmes foctal metabolism and cardiovascular function for the development of obesity and its related cardiovascular diseases in adult life. The hypolipidemic actions of coconut water (CW) in the presence of HFD have been reported. This study examined the effects of oral administration of CW on lipid panel, hormone profile, pup and placental morphometry of dams fed HFD during gestation.

Methods: Twenty-four pregnant Wistar rats were assigned to four groups (n=6) and treated daily from gestation day (GD) 1 to 21 as follows; Group 1: 1ml/ 100g b.wt. distilled water; Group 2: 1ml/100g b.wt. CW; Group 3: HFD (70% standard rat feed plus 30% butter); Group 4: HFD + 1ml/100g b.wt. CW. Animals were sacrificed on GD 21. Random blood glucose was measured using tail blood. Caesarean section was performed to remove the pups and their placentas which were immediately measured. Oxidative stress status of the placentas; serum lipid and hormone profiles of dams were assessed.

Results: HFD+CW resulted in significant (P<0.05) reductions in pup weight and morphometric indices when compared with pups from HFD. These changes were accompanied by significant improvements in maternal serum lipid profile, alterations in hormone levels and higher placental lipid peroxidation.

Conclusion: These results suggest that coconut water is protective against maternal high fat diet-induced changes. Further studies are on-going to determine the actions of coconut water of maternal high fat diet induced foetal programming of adult health.

Keywords; Maternal high fat diet: coconut water; morphometry

Résumé

Contexte: Le régime maternel riche en haute graisse (RRHG) pendant la gestation défavorablement programme le métabolisme fœtal et la fonction

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cardiovasculaire pour le développement de l'obésité et ses maladies cardiovasculaires liées dans l'âge adulte. Les actions hypo lipidiques de l'eau de noix de coco (EC) en présence de RRHG ont été rapportées. Cette étude a examiné les effets de l'administration orale d'EC sur panneau lipidique, profil hormonal, souriceaux et morphométrie placentaire de barrages nourris avec RRHG pendant la gestation.

Méthodes: Vingt-quatre rats Wistar gestantes ont été réparties en quatre groupes (n = 6) et traitées tous les jours depuis le jour de gestation (JG) 1 à 21 comme suit; Groupe 1: 1 ml / 100 g de poids corporel eau distillée; Groupe 2: 1 ml / 100 g de poids corporel EC; Groupe 3: RRHG (70% d'alimentation standard de rat plus 30% de beurre); Groupe 4: RRHG + 1 ml / 100g de poids corporel EC. Les animaux ont été sacrifiés le JG 21. Glucose de sang aléatoire a été mesuré en utilisant le sang provenant de la queue. La césarienne a été effectuée pour enlever les souriceaux et leur placenta qui ont été immédiatement mesurées. L'état de stress oxydatif des placentas; lipide sérique et profils hormonaux de barrages ont été évalués.

Résultats: RRHG + EC ont entraîné des réductions significatives (P <0,05) du poids des souriceaux et indices morpho-métrique par rapport aux souriceaux de RRHG Ces changements ont été accompagnés par des améliorations significatives en profil maternel des lipides sériques, des altérations dans les niveaux d'hormones et plus haute peroxydation de lipide placentaire.

Conclusion: Ces résultats suggèrent que l'eau de coco est protectrice contre les changements induits par le régime riche en graisses maternels. D'autres études sont en cours pour déterminer les actions de l'eau de noix de coco de régime maternel riche en graisse de programmation fœtal induit de la santé d'adulte.

Mots-clés; Régime maternel riche en graisses; eau de noix de coco; morphométrie

Introduction

Obesity is a global health problem; its negative impact cuts across socioeconomic class and national development. Although the actiology of obesity is complex and probably not fully understood, it has been known to involve the integration of social, behavioural, cultural, physiological, metabolic and genetic factors [1]. Maternal exposure to any of these



factors can program future generations of offspring to develop obesity irrespective of the offspring's exposure [2,3]. The foctal programming hypothesis proposes that modifications in maternal nutrition and endocrine status could result in developmental adaptations of the foctus which permanently alter its structure, physiology and metabolism [4]. This has since been proven by several epidemiological studies and also through human and animal experimental models [5-8]. Morphometric indices such as body weight, height, head and abdominal circumferences, are very useful in assessing foctal/ neonatal growth and development [9]. For instance, a u-shaped curve association between the risk of adult disease and birth weight has been well established [10]. These measures are often standardized as ratios such as the Ponderal Index (PI), waist-to-height ratio (WHtR), waist-to-hip ratio (WHR) and head circumference-to-abdominal circumference ratio (HC/AC); thereby enabling comparison between populations and discoveries of associations between body size and disease risk [11-14].

Maternal obesity or consumption of high caloric dicts during pregnancy, leads to foctal programming of offspring, thereby predisposing them to the development of obesity along with its related complications in later life [3,6,15-17]. The deleterious effects of maternal high fat diet on the neuroendocrine, metabolic, cardiovascular and reproductive health of offspring have been widely reported [18-31]. Therefore, it stands to reason that campaigns for healthy maternal nutrition should be intensified. However, the prevailing socioeconomic circumstances in developing countries inadvertently sabotage the efficacy of such campaigns. The population of females in the work force has remarkably increased since the 1960 to 1980 era when women first joined the labour market, with some countries recently recording over 80% female participation [32,33]. This implies that the only diet options for several mothers during the perinatal period are the readily available, energy dense, high fat containing, fast foods [34]. Feasible solutions to maternal high fat diet induced foetal programming should therefore incorporate more convenient alternatives in addition to dietary modifications. Studies suggest that such adverse programming of offspring can be potentially reversed by nutritional or targeted therapeutic interventions especially during the period of developmental plasticity [35].

Some of the reactions proposed for the adverse perinatal effects of high maternal fat diet include oxidative stress, lipotoxicity and inflammation [31,36–38]. The hypolipidemic, hypoglycaemic and antioxidant effects of coconut water, the liquid endosperm of the coconut (*Cocos nuciferaL.*) fruit, have been reported [39–42].

Coconut water has no reported toxicity and can hence be considered as a safe and convenient option for career mothers. This study was therefore designed to investigate the effects of coconut water administration and maternal high fat diet during gestation on maternal lipid profile and pup morphometry in Wistar rats.

Materials and methods

Plant material

Coconut (*Cocos mucifera* L.)fruits were obtained from a coconut plantation in Oyo state, Nigeria and verified by a botanist from the Department of Botany, University of Ibadan, Ibadan, Nigeria. Coconut water was obtained by piercing the soft "eye" of the coconut (the germination pore) with a sterile screw driver and decanting the water into a large sterile container. Fresh coconut water was used each day. Coconut water was administered via oral gavage at a daily dosage of 1 ml/ 100g body weight using blunt-tipped oral cannulas attached to 2ml syringes. Administration of coconut water was done between 8.00-9.00am daily.

Animals

All procedures involving animals in this study conformed to the guiding principles for research involving animals as recommended by the guidelines for laboratory animal care of the National Institute of Health (NIH publication no. 85-23, revised 1996). Virgin female rats obtained from the Central Animal House, University of Ibadan were mated with proven breeder male rats from the Laboratory for Reproductive Physiology and Developmental Programming, Department of Physiology, University of Ibadan. Pregnancy was confirmed by the presence of spermatozoa in vaginal smears and the day of observation of spermatozoa was taken as gestation day (GD) 1 for each female. Pregnant rats were then randomly divided into four groups, namely; control, coconut water (CW), high fat diet (HFD) and high fat diet plus coconut water (HFD+CW).

Treatments

High fat diet consisted of 70% standard rodent diet (Ladokun Feed Mills, Ibadan, Nigeria) and 30% butter (Real brand, Chellarams, Lagos, Nigeria). With the exception of the HFD and HFD+CW groups which received high fat diet during gestation, all rats were fed standard rodent diet before mating and during gestation. The animals had access to food and drinking water *ad libitum*.

Caesarean section

On the twenty-first day of gestation (GD21), ether anaesthesia was induced by placing the rat in an airtight desiccator containing a ball of cotton wool

moistened with a few drops of diethyl ether for about one minute until the evelid closure reflex was lost. A drop of tail blood was collected for random blood glucose measurements which were done using an

automated glucometer (On Call Plus®, ACON Laboratories Inc., USA). Respiratory movements were monitored visually by observing for regular chest and abdominal undulations. The anaesthetized dams were cut open from the linea alba of the anterior abdominal wall to the thoracic cavity to expose the heart. Blood was collected via cardiac puncture using sterile needles and syringes and emptied into plain tubes. The abdominal cavity was then dissected to remove the gravid uterus.

Pup and placental morphometry

The pups and their placentas were immediately removed from within the uteri and weighed individually on an electronic scale (Lisay, China). Crown-to-rump length, abdominal diameter and head diameter of each pup; diameter and thickness of each placenta were measured using a digital Vernier calliper (Mitutoyo, Japan). The diameter was measured along the length of each placenta, while the thickness was measured at the centre when the placenta was placed on a horizontal plane.

Placenta redox status

The largest placenta from each dam was homogenized in 4ml of phosphate buffer (pH 7.4) per gram of placental tissue. The supernatant was obtained after centrifuging at 3000rpm for 15 minutes and was used for the determination of malondialdehyde (MDA), superoxide dismutase

Hormone assays

Serum collected was used to assay for follicle stimulating hormone (FSH), luteinizing hormone (LH), oestrogen, testosterone (Fortress Diagnostics Limited, UK), corticosterone (Oxford Biomedical Research, USA), insulin and leptin (Ray Biotech Inc. USA) using the ELISA technique. They were determined using kits according to the manufacturer's instructions.

Statistics

Data are expressed as Mean ± Standard Error of Mean (SEM). Significance of difference of means was analysed using one-way ANOVA followed by post hoc analysis where necessary. P<0.05 was considered significant.

Results

Body weight, serum lipid and blood glucose levels of dams at GD21

High fat diet led to a significant gain in body weight during gestation when compared to the control and coconut water (CW) groups (Fig.1). High fat diet (HFD)dams showed a statistically significant (p<0.05) increase in serum triglyceride, total cholesterol and low density lipoprotein (LDL) cholesterol concentrations during gestation (Fig. 2) which was not evident in HFD+CW dams. Maternal high density lipoprotein (HDL) cholesterol was reduced in all the groups when compared with control (Fig.2). Leptin secretion was significantly increased in HFD dams and reduced in HFD+CWdams (Fig.3). CW dams showed a significant reduction in random blood glucose levels (Fig. 4), while serum insulin levels were significantly



Fig.1: Body weight gain of dams during gestation. Initial body weight was measured on GD1 while final body weight was measured on GD21. P<0.05 was considered significant when compared with "control and "CW groups respectively.

(SOD), catalase and glutathione peroxidase (GPx) levels using standard assay techniques [43-45].

increased in the HFD dams when compared with CW dams (Fig.5)



Fig.2: Serum lipid profile of dams on GD21. P<0.05 was considered significant when compared with *control,*CW and *HFD+CW groups respectively



Fig.3: Serum leptin levels of dams on GD 21. P<0.05 was considered significant when compared with ^acontrol,^bCW and ^cHFD+CW groups respectively.



Fig.4: Random blood glucose concentration of dams on GD 21. P<0.05 was considered significant when compared with *control and *CW groups respectively.



Fig.5: Serum insulin concentration of dams on GD 21. P<0.05 was considered significant when compared with CW group.



Fig.6. Serum levels of the gonadotropins; Luteinizing hormone (LH) and Follicle Stimulating hormone (FSH) on GD 21. P<0.05 was considered significant when compared with *control, *CW and *HFD+CW groups respectively.



Fig.7: Serum levels of the steroid hormones; Estradiol, Testosterone and Corticosterone, on GD 21. P<0.05 was considered significant when compared with "control and "CW groups respectively.

Hormone profile of dams on GD21

Scrum Luteinizing hormone (LH) levels were significantly increased in CW and reduced in HFD dams (Fig.6). Scrum Follicle Stimulating hormone (FSH) levels were significantly increased in HFD+CW dams when compared with CW dams (Figure 6). Serum levels of estradiol on GD21 were significantly increased in all the test groups (Fig.7).



 CONTROL
 CW
 HFD
 HFD+CW

 Fig.8: Weight of pups on GD 21. P<0.05 was considered significant when compared with "control and"CW groups respectively.</td>







Fig.10: Ponderal index (PI) of offspring on PND 1. P<0.05 was considered significant when compared with 'control, ^bCW and ^cHFD groups respectively.

Serum testosterone levels were increased in HFD+CW dams when compared with the controls (Fig.7). Compared with the CW dams, HFD dams showed a decrease in serum testosterone levels (Fig. 7).Serum corticosterone levels were not significantly affected in this study (Fig.7).

Pup morphometry

There was a statistically significant decrease in the weight of pups from all the groups on GD21, with coconut water offspring showing the most pronounced reduction (Fig.8). The head circumference of offspring on PND 1 was



Fig.11: Waist-to-height and Head-to-abdomen ratios of pups on PND 1. P<0.05 was considered significant when compared with *control, ^bCW and 'HFD groups respectively.



Fig.12: Placental weight on GD21.P<0.05 was considered significant when compared with "control and "CWgroups respectively.



Fig. 13: Placental thickness and diameter on GD21. P<0.05 was considered significant when compared with *control, *CW and *IIFD groups respectively

significantly increased by maternal CW administration and reduced by HFD+CW, while the abdominal circumference was increased in both CW and HFD (Fig.9). The crown-to-rump length

(height) of pups was significantly reduced in HFD+CW (Fig.9). Ponderal index (PI) at birth was significantly increased in HFD offspring, while the CW and HFD+CW offspring showed a reduction in PI (Fig.10). An increase in waist-to-height ratio and a reduction in head-to-abdomen ratio were observed in CW and HFD offspring; no significant difference was however observed for both variables in HFD+CW offspring (Fig.11).

Placental morphometry and redox status

The weight and size of the placenta was significantly reduced in CW group (Figs.12 and 13). There was significantly more lipid peroxidation in both the CW and HFD+CW groups as indicated by the increased malondialdehyde concentration (Table 1). However, this process did not adversely affect the concentration of antioxidants in the placenta of these groups on GD 21 (Table 1). Placentas from the HFD group had significantly higher glutathione peroxidase and significantly lower superoxide dismutase concentrations than the control placentas (Table 1). resultant health complications [51]. This occurs as a result of an increase in neurogenesis in the hypothalamic third ventricle and the increased expression of orexigenic peptides in the developing foctus [51]. Maternal HFD is thus a major cause for concern as dietary-induced obesity is becoming more prevalent among women than men in all regions of the world [52]. Hence, multi-interventional approaches are required to protect future generations from an impending obesity epidemic.

Coconut water is a natural, pleasant-tasting and sterile drink which contains several biologically active compounds which include; L-arginine, ascorbic acid, calcium, magnesium and potassium [41,53,54]. Numerous health benefits have been ascribed to coconut water [42,53,55,56]. The antioxidant, antidiabetic and hypolipidemic effects

Table 1: Placental malondialdehyde and antioxidant levels on GD 21.

Groups	MDA (μmol/g tissue)	CATALASE (U/g tissue)	SOD (U/g tissue)	GPx (U/g tissue)
Control	1.35±0.07	23.30±1.72	3.47±0.20	1.78±0.12
Coconut Water	2.34±0.41*	27.73±4.14	2.65 ± 0.24	2.24±0.42
High Fat Dict	1.52 ± 0.12	19.98±0.38 ^b	2.09±0.36*	3.95±0.10 ^{ab}
High Fat Dict +	2			
Coconut Water	2.62±0.42 ^{±c}	22.96±2.43	3.08±0.61	2.46±0.47°

MDA=malondialdehyde, SOD=superoxide dismutase, GPx=glutathione peroxidase. P<0.05 was considered significant when compared with *control, *CW and *HFD groups respectively.

Discussion

Foctal programming is a concept which refers to how maternal nutrition and environmental exposures result into in utero alterations in foetal structure and function which permanently modify adult physiology of offspring [4,46]. Maternal obesity and high fat diet during gestation programme offspring for the development of obesity, metabolic syndrome and cardiovascular dysfunction among other related diseases [16,22,47,48]. Pregnancy is normally associated with a gain in body weight which could be associated with the growing foctus(es) and/or an increase in maternal food intake to cater for the increased metabolic requirements. A high fat diet (HFD) during gestation upsurges the pregnancyinduced weight gain and portends a negative impact on the future health of the unborn offspring [48-50]. Maternal HFD during gestation programmes the offspring with alterations which predispose them to overweight, hyperphagia (along with a preference for fat) and hyperlipidaemia together with the

may hold promise for the development of sustainable therapies in metabolic dysfunction.

The hypolipidemic effect of coconut water reflected in this study was observed only when the concentration of dietary fats was increased. This supports previous findings [40]. The results of this study also suggest that coconut water potentiates the actions of leptin when there is an abundance of circulating fat, without affecting leptin synthesis. Leptin is an adipokine which regulates maternal to foctal metabolic interactions during pregnancy [57]. The mechanism via which coconut water alters the interplay of leptin, insulin and energy balance requires further investigation as these results suggest the absence of central leptin resistance and significant insulin levels (which are normal features in late gestation) [58]. The sex hormone activity on serum leptin levels is also contrary to what was reported by Ahima and Flier [59] that testosterone decreases and oestrogen increases leptin levels, thereby suggesting an independent action of coconut water on lipid metabolism. These results also imply

environment. Further studies are on-going to establish the foctal programming outcomes of the adult offspring since the foctal programming hypothesis relates to health of offspring in later life [35,46,68].

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References

- Al-Quaiz AJ. Current concepts in the management of obesity. An evidence based review. Saudi Med J. 2001. 22(3):205–210.
- Heerwagen MJR, Miller MR, Barbour LA and Friedman JE. Maternal obesity and fetal metabolic programming: a fertile epigenetic soil. Am J Physiol Regul Integr Comp Physiol. 2010. 299(3):R711–R722
- Tenenbaum-Gavish K and Hod M. Impact of maternal obesity on fetal health. Fetal Diagn Ther. 2013. 34(1):1-7.
- 4. Barker DJP. The fetal and infant origins of disease. Eur J Clin Invest. 1995. 25(7):457-463.
- Godfrey K and, Barker DJP. Fetal nutrition and adult disease. Am J Clin Nutr. 2000. 71(5):1344S – 1352S.
- Armitage JA, Taylor PD and Poston L. Experimental models of developmental programming: consequences of exposure to an energy rich diet during development. J Physiol. 2005. 565(Pt 1):3-8.
- Drever N, Saade GR and Bytautiene E. Fetal programming: Early-life modulations that affect adult outcomes. Current Allergy and Asthma Reports. 2010. p. 453–459.
- Sookoian S, Gianotti TF, Burgueño AL and Pirola
 CJ. Fetal metabolic programming and epigenetic modifications: a systems biology approach.
 Pediatr Res. 2013. 73(4 Pt 2):531–542.
- Guihard-Costa AM and Larroche JC. Growth velocity of some fetal parameters. II. Body weight, body length and head circumference. Biol Neonate. 1992. 62(5):317–324.
- 10. Thompson LP and Al-Hasan Y. Impact of oxidative stress in fetal programming. J Pregnancy. 2012. (Article ID 582748):8 pages.
- Barker DJP, Godfrey KM, Osmond C and Bull A. The relation of fetal length, ponderal index and head circumference to blood pressure and the risk of hypertension in adult life. Paediatr Perinat Epidemiol. 1992. 6(1):35-44.

the rate were given coconut water only once a day ine rate were given coconut water only once a day and at a dose much lower than what had been previously reported [39,40]. Sandhya and Kajamohan [39,40] have proposed several mechanisms of action for the hypolipidemic effects of coconut water in the presence of dietary fats. Notable among these are the enhanced hepatic lipid metabolism reflected by an increase in the activity of 3-hydroxy-3-methylglutaryl-CoA reductase(which catalyses the rate-limiting step in cholesterolsynthesis) and a reduction in lipogenic enzymeactivity along with an increased conversion ofactivity along with an increased conversion ofactivity along with an increased conversion ofavailable cholesterol to bile acids and an increasedavailable cholesterol to bile acids and an increasedavailable cholesterol to bile acids and an increasedexerction of these bile acids [<math>39,40,60].

synthesis. oxide which in turn stimulates LH and FSH coconut water stimulate the production of nitric proposed that the L-arginine and ascorbic acid in coconut water [64]. Nair and Rajamohan [54] reports of a hypothalamo-pituitary axis activity of concentration observed in CW supports previous a negative feedback theory. The increase in LH concomitant reduction in serum LH levels supports of increased availability of precursors and the be due to an increase in steroidogenesis as a result increase in estradiol level observed with HFD may natural ocstrogen replacement therapy [63]. The water [62]. Phytoestrogens have been promoted as be due to the presence of phytoestrogens in coconut The increase in serum estradiol observed in CW may observed in this study further supports that fact [61]. of coconut water and the zero maternal mortality No toxicity has been reported with the use

hypolipidemic effects may adversely alter the foetal temales on a healthy balanced dict as its coconnt water should not be consumed by pregnant induced foctal changes during gestation, and that water may protect against maternal high fat diet [12,67]. It was therefore concluded that coconut inclination to disease development in later life parts have been suggested to act as pointers of high birth weight and a relative disproportion of body maternal diet. Morphometric indices such as low/ detrimental when excess fat is not available in presence of maternal high fat diet and may be water has a protective effect on the pups in the in the HFD+CW offspring suggests that coconut significant differences in pup morphometry observed compromises maternal health [65,66]. However, the reduces placental efficiency and thereby coconut water intake suggests that coconut water peroxidation in the placenta caused by maternal The reduction in size and increased lipid

- Barker DJP, Osmond C, Simmonds SJ and Wield GA. The relation of small head circumference and thinness at birth to death from cardiovascular disease in adult life. BMJ. 1993. 306(6875):422– 426.
- Esmaillzadeh A, Mirmiran P and Azizi F. Waistto-hip ratio is a better screening measure for cardiovascular risk factors than other anthropometric indicators in Tehranian adult men. Int J Obes Relat Metab Disord. 2004. 28(10):1325–1332.
- Lee KS, Eom JS, Cheong HK, *et al.* Effects of head circumference and metabolic syndrome on cognitive decline. Gerontology. 2010. 56(1):32– 38.
- Desai M, Beall M and Ross MG. Developmental origins of obesity: programmed adipogenesis. Curr Diab Rep. 2013. 13(1):27–33.
- Burgueño AL, Cabrerizo R, Gonzales Mansilla N, et al. Maternal high-fat intake during pregnancy programs metabolic-syndromerelated phenotypes through liver mitochondrial DNA copy number and transcriptional activity of liver PPARGC1A. J Nutr Biochem. 2013. 24(1):6–13.
- 17.Srinivasan M, Dodds C, Ghanim H, et al. Maternal obesity and fetal programming : effects of a high-carbohydrate nutritional modification in the immediate postnatal life of female rats. Am J Physiol Endocrinol Metab. 2008. 295:E895-903.
- Khan I, Dekou V, Hanson M, Poston L and Taylor P. Predictive adaptive responses to maternal high-fat diet prevent endothelial dysfunction but not hypertension in adult rat offspring. Circulation. 2004. 110(9):1097-1102.
- Khan IY, Dekou V, Douglas G, et al. A high-fat diet during rat pregnancy or suckling induces cardiovascular dysfunction in adult offspring. Am J Physiol Regul Integr Comp Physiol. 2005. 288(1):R127–R133.
- Cerf ME, Williams K, Nkomo XI, et al. Islet cell response in the neonatal rat after exposure to a high-fat diet during pregnancy. Am J Physiol Regul Integr Comp Physiol. 2005. 288(5):R1122–R1128.
- Srinivasan M, Katewa SD, Palaniyappan A, et al. Maternal high-fat diet consumption results in fetal malprogramming predisposing to the onset of metabolic syndrome-like phenotype in adulthood. Am J Physiol Endocrinol Metab. 2006. 291(4):E792–E799
- 22. Férézou-Viala J, Roy A-F, Sérougne C, et al. Long-term consequences of maternal high-fat

feeding on hypothalamic leptin sensitivity and diet-induced obesity in the offspring. Am J Physiol Regul Integr Comp Physiol. 2007. 293(3):R1056-R1062

- 23. Naef L, Srivastava L, Gratton A, et al. Maternal high fat diet during the perinatal period alters mesocorticolimbic dopamine in the adult rat offspring: Reduction in the behavioural responses to repeated amphetamine administration. Psychopharmacology. 2008. 197(1):83–94.
- Bruce KD, Cagampang FR, Argenton M, et al. Maternal high-fat feeding primes steatohepatitis in adult mice offspring, involving mitochondrial dysfunction and altered lipogenesis gene expression. Hepatology. 2009. 50(6):1796–1808.
- 25. Naef L, Moquin L, Dal Bo G et al. Maternal high-fat intake alters presynaptic regulation of dopamine in the nucleus accumbens and increases motivation for fat rewards in the offspring. Neuroscience. 2011. 176:225–236.
- 26. Sullivan EL, Smith MS and Grove KL. Perinatal exposure to high-fat diet programs energy balance, metabolism and behavior in adulthood. Neuroendocrinology. 2011. 93(1):1–8
- Ashino NG, Saito KN, Souza FD, *et al.* Maternal high-fat feeding through pregnancy and lactation predisposes mouse offspring to molecular insulin resistance and fatty liver. J Nutr Biochem. 2012. 23(4):341–348.
- Volpato AM, Schultz A, Magalhães-da-Costa E, et al. Maternal high-fat diet programs for metabolic disturbances in offspring despite leptin sensitivity. Neuroendocrinology. 2012. 96(4):272–284.
- Peleg-Raibstein D, Luca E and Wolfrum C. Maternal high-fat diet in mice programs emotional behavior in adulthood. Behav Brain Res. 2012. 233(2):398–404.
- Benesh EC, Humphrey PA, Wang Q and Moley KH. Maternal high-fat diet induces hyperproliferation and alters Pten/Akt signaling in prostates of offspring. Sci Rep.2013. 3:3466
- Sullivan EL, Nousen L, Chamlou KKA and Nousen EK. Maternal high fat diet consumption during the perinatal period programs offspring behavior. Physiol Behav. 2014. 123:236–242.
- Kaestner R. Obesity/ : Causes, Consequences and Public Policy Solutions. The Illinois Report. 2009.
- 33. The World Bank. Labor force participation rate, female (% of female population ages 15+)(modeled ILO estimate). World Development Indicators. 2014. http:// data.worldbank.org/indicator/SL.TLF.CACT.ZS

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- Currie J, DellaVigna S, Moretti E and Pathania V. The effect of fastfood restaurants on obesity and weight gain.NBER Working Paper 14721. 2009. http://www.nber.org/papers/w14721
- Vickers MH. Developmental programming of the metabolic syndrome - critical windows for intervention. World Journal of Diabetes. 2011. p. 137.
- McCurdy CE, Bishop JM, Williams SM, *et al.* Maternal high-fat diet triggers lipotoxicity in the fetal livers of nonhuman primates. J Clin Invest. 2009. 119(2):323–235.
- Dong M, Zheng Q, Ford SP, *et al*.Nathanielsz PW, Ren J. Maternal obesity, lipotoxicity and cardiovascular diseases in offspring. J Mol Cell Cardiol. 2013. 55(1):111–116.
- 38. Yokomizo H, Inoguchi T, Sonoda N, et al. Maternal high fat diet during gestation and lactation affects glucose metabolism and pancreatic (beta) cell function in mature offspring in mice. Diabetes. 2011. 60:A532.
- Sandhya VG and Rajamohan T. Beneficial effects of coconut water feeding on lipid metabolism in cholesterol-fed rats. J Med Food. 2006. 9(3):400–407.
- Sandhya VG and Rajamohan T. Comparative evaluation of the hypolipidemic effects of coconut water and lovastatin in rats fed fatcholesterol enriched diet. Food Chem Toxicol. 2008. 46(12):3586–3592.
- Bhagya D, Prema L and Rajamohan T. Beneficial effects of tender coconut water on blood pressure and lipid levels in experimental hypertension. J cell tissue Res. 2010. 10(1):2139–2144.
- 42. Preetha PP, Devi VG and Rajamohan T. Antihyperlipidemic effects of mature coconutwater and its role in regulating lipid metabolism in alloxan-induced experimental diabetes. Comp Clin Path. 2014. 23(5):1331–1337.
- Misra HP and Fridovich I. Superoxide dismutase: A photochemical augmentation assay. Arch Biochem Biophys. 181(1):308–312.
- Sinha AK. Colorimetric assay of catalase. Anal Biochem. 1972. 47(2):389–394.
- Rotruck JT, Pope AL, Ganther HE, et al. Selenium: Biochemical Role as a Component of Glutathione Peroxidase. Science. 1973. 179(4073):588–590.
- Godfrey KM and Barker DJP. Fetal programming and adult health. Public Health Nutr. 2007. 4(2B):611–624.
- 47. Shankar K, Harrell A, Liu X, et al. Maternal obesity at conception programs obesity in the

offspring. Am J Physiol Regul Integr Comp Physiol. 2008. 294(2):R528-R538.

- Guberman C, Jellyman JK, Han G, Ross MG and Desai M. Maternal high-fat diet programs rat offspring hypertension and activates the adipose renin-angiotensin system. Am J Obstet Gynecol. 2013. 209(3):262.e1–8.
- Kamimac-Lanning AN, Krasnow SM, Goloviznina NA, et al. Maternal high-fat diet and obesity compromise fetal hematopoiesis. Mol Metab. 2015. 4(1):25–38.
- 50. Franco JG, Fernandes TP, Rocha CPD, et al. Maternal high-fat diet induces obesity and adrenal and thyroid dysfunction in male rat offspring at weaning. J Physiol. 2012. 590(Pt 21):5503-5518.
- 51. Chang GQ, Gaysinskaya V, Karatayev O and Leibowitz SF. Maternal high-fat diet and fetal programming: increased proliferation of hypothalamic peptide-producing neurons that increase risk for overcating and obesity. J Neurosci. 2008. 28(46):12107–12119.
- 52. World Health Organisation Global Health Observatory data. Obesity Situation and Trends. http://www.who.int/gho/ncd/risk_factors/ obesity_text/en/#. 2014.
- 53. Preetha PP, Devi VG and Rajamohan T. Hypoglycemic and antioxidant potential of coconut water in experimental diabetes. Food Funct. 2012. 3(7):753.
- 54. Nair SVG and Rajamohan T. The Role of Coconut Water on Nicotine-Induced Reproductive Dysfunction in Experimental Male Rat Model. Food Nutr Sci. 2014. 05(12):1121–1130
- 55. DebMandal M and Mandal S. Coconut (Cocos nucifera L.: Arecaceae): In health promotion and disease prevention. Asian Pac J Trop Med.
 2011;4(3):241-247.
 - Prathapan A and Rajamohan T. Antioxidant and antithrombotic activity of tender coconut water in experimental myocardial infarction. J Food Biochem. 2011. 35(5):1501–1507.
 - Margetic S, Gazzola C, Pegg G and Hill R. Leptin: a review of its peripheral actions and interactions. Int J Obes Relat Metab Disord. 2002. 26(11):1407–1433.
 - Tessier D, Ferraro Z and Gruslin A. Role of leptin in pregnancy: consequences of maternal obesity. Placenta. 2013. 34(3):205–211.
 - Se Ahima R and Flier J. Leptin. Annu Rev Physiol. 2000. 62(1):413–437.
 - Friesen J and Rodwell V. The 3-hydroxy-3methylglutaryl coenzyme-A (HMG-CoA) reductases. Genome Biol. 2004. 5(11):248.

- Eiseman B, Lozano R and Hager T. Clinical experience in intravenous administration of coconut water. AMA Arch Surg. 1954. 69(1):87– 93.
- 62. Radenahmad N, Saleh F, Sawangjaroen K, et al. Young coconut juice, a potential therapeutic agent that could significantly reduce some pathologies associated with Alzheimer's disease: novel findings. Br J Nutr. 2011. 105(5):738–746.
- Patisaul H and Jefferson W. The pros and cons of phytoestrogens. Front Neuroendocr. 2010. 31(4):400–419.
- Kunle-Alabi OT, Akindele OO, Oyovwi MO, et al. Cocos nucifera L. water improves reproductive indices in Wistar Rats. Afr J Med med Sci. 2014. 43:305–313.

- 65. Madazli R, Benian A, Aydin S, et al. The plasma and placental levels of malondialdehyde, glutathione and superoxide dismutase in precelampsia. J Obs Gynaecol. 2002. 22(5):477–480
- 66. Madazli R, Tuten A, Calay Z, *et al.* The incidence of placental abnormalities, maternal and cord plasma malondialdehyde and vascular endothelial growth factor levels in women with gestational diabetes mellitus and nondiabetic controls. Gynecol Obs Invest. 2008. 65(4):227–232.
- Robinson R. The fetal origins of adult disease: no longer just a hypothesis and may be critically important in south Asia. BMJ.2001. 41(6):158– 176.
- Gicquel C, El-Osta A and Le Bouc Y. Epigenetic regulation and fetal programming. Best Pract Res Clin Endocrinol Metab. 2008. 22(1):1–16.

